A population dynamics model of Streptococcus pneumoniae using genomics & Implementing a model translation and visualisation tool

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A population dynamics model of *Streptococcus pneumoniae* using genomics

Streptococcus pneumoniae



- Serotypically diverse bacteria with more than 100 serotypes
- Genetically diverse with hundreds of lineages/strains
- Major cause of diseases, including pneumonia and meningitis, especially in young children under 2 years of age and adults over 65 years of age
- Treatment: antibiotics
- Prevention: pneumococcal vaccines



Pneumococcal vaccines





Pneumococcal vaccines



Simple compartmental Model





Compartmental Model with Sub-compartments



7

Data

Parameters: Compartmental Model with NFDS σ_f **Sub-compartments prop**_f vaccination v migration *m* $P_{i,j}^{t+1} = B_{i,j}^t + M_{i,j}^t$ **Births** Population Migration $B_{i,j}^t \sim Poisson(K \cdot (\frac{p_{i,j}^{\iota}}{\sum_{i=1}^n \sum_{i=1}^l p_{i,i}^t}) * (1 - \mathbf{m}))$ $M_{i,j}^t \sim binomial(\hat{m}^t, m_{i,j})$ $\hat{m}^t \sim \text{binomial}(K, \mathbf{m})$ $p_{i,j}^{t} = (1 + \sigma_{\mathbf{f}})^{\pi_{i,t}} * P_{i,j}^{t} * (1 - (v_{\text{time}}(t) * v_{\text{type}}(j) * \mathbf{v}))$ $\sum e_l - f_{l,t}$ $\pi_{i,t} =$

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NFDS = Negative frequency-dependent selection,

Model based on Jukka Corander et al. "Frequency-dependent selection in vaccine-associated pneumococcal population dynamics". (2017)

```
43 # identify genes that are under NFDS
44 pi_f_freq[,] <- if ((delta[i] <= prop_f * gene_no)) Genotypes[i,j] * (eq[i] - freq[i]) else 0
45 pi_f_genotypes[] <- sum(pi_f_freq[1:gene_no,i])
46
47 # Genotype specific probability to produce offspring
48 # those are the individuals' probabilities multiplied by the number of individual that have this genotype
49 probs[,] <- (1 + exp(sigma_f))^pi_f_genotypes[i] * Pop[i,j] * (1- (as.integer(time >= vacc_time) * vaccTypes[j] * v))
50
51 # generate the next generation based on the current one
52 y[,] <- if ((probs[i,j]/sum(probs[1:species_no,1:sero_no])) < 1)
     rpois(capacity * (probs[i,j] / sum(probs[1:species_no,1:sero_no])) * (1-exp(m)) ) else rpois(capacity * 1 *(1-exp(m)) )
53
54
55 # m is the miaration rate
56 # fitness of individuals in the community is reduced by this rate
57 # determining migration number:
58 mia_num <- rbinom(capacity, exp(m))</pre>
59 Pop_mig[,] <- rbinom(mig_num, migVec[i,j])</pre>
60
61 ## Core equation for population (assume constant size here):
62 update(Pop[,]) <- y[i,j] + Pop_mig[i,j]
63 update(Pop_tot[]) <- sum(y[i,]) + sum(Pop_mig[i,])
64
65 initial(Pop[,]) <- Pop_ini[i,j] # deterministic, user-based start value
66 initial(Pop_tot[]) <- sum(Pop_ini[i,])
67
```



Strain-serotype Model Fit to Massachusetts





Strain-serotype Model Fit to Massachusetts



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Strain-serotype Model Fit - Parameter Comparison

Parameter	Massachusetts	Southampton	Nepal
neg. log likelihood	-236.12234	-562.76	-2415.2916
	(-239.9329 , -233.6487)	(-567.6667, -560.4743)	(-2419.1133, -2412.7752)
Strength of NFDS $\sigma_{\rm f}$	0.0345	0.0379	0.4670
	(0.0144, 0.0685)	(0.0133, 0.0826)	(0.3474, 0.5386)
Proportion of genes	0.28835	0.1614	0.1708
under NFDS prop_f	(0.20115, 0.4566)	(0.1077, 0.3611)	(0.1584, 0.1967)
Migration rate m	0.0133	0.0185	0.0792
	(0.0087, 0.0214)	(0.0126, 0.0252)	(0.0648, 0.0953)
Vaccination selection strength v	0.0814	0.0905	0.1766
	(0.0627, 0.1046)	(0.0759, 0.1055)	(0.1405, 0.2159)

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Benefits of implementing this model in odin

- Defining variables as vectors / matrices
- Readability of the model code (looks like equations)
- Reproducibility when fitting the model
- Having a likelihood and using MCMC
- It's fast

--> Reusability



Implementing a model translation and visualisation tool

Model Accessibility





Model Accessibility

Elowitz2000 - Repressilator				
View the 2006-07 Model of the Month entry for this model >				
Overview Files History	Components Curation			
Model Identifier Short description	BIOMD000000012 Elowitz2000 - Repressilator This model describes the deterministic version of the repressilator system.	Metadata information		
	The authors of this model (see reference) use three transcriptional repressor systems that are not part of any natural biological clock to build an oscillating network that they called the repressilator. The model system was induced in Escherichia coli.	Js (2 statements) BIOMOdels Database BIOMD0000000012 BIOModels Database MODEL6615351360		
	In this system, LacI (variable X is the mRNA, variable PX is the protein) inhibits the tetracycline-resistance transposon tetR (Y, PY describe mRNA and protein). Protein tetR inhibits the gene CI from phage Lambda (Z, PZ: mRNA, protein), and protein CI inhibits lacI expression. With the appropriate parameter values this system oscillates.	JsDescribedBy (1 statement) PubMed 10659856		
	This model is described in the article:	hasTaxon (1 statement) Taxonomy Escherichia coli		
	A synthetic oscillatory network of transcriptional regulators. Elowitz MB, Leibler S. Nature. 2000 Jan; 403(6767):335-338 Abstract:	JSVRrSignQf (1 statement) Gene Ontology regulation of gene expression, epigenetic		
	Networks of interacting biomolecules carry out many essential functions in living cells, but the 'design principles' underlying the functioning of such intracellular networks remain poorly understood, despite intensive efforts including quantitative analysis of relatively simple systems. Here we present a complementary approach to this problem: the design and construction of a synthetic network to implement a particular function. We used three transcriptional	Mathematical Modelling Ontology Ordinary differential equation model		
	repressor systems that are not part of any natural biological clock to build an oscillating network, termed the repressilator, in Escherichia coll. The network periodically induces the synthesis of green fluorescent protein as a readout of its state in individual cells. The resulting oscillations, with typical periods of hours, are slower than the cell-division cycle, so the state of the oscillator has to be transmitted from generation to generation. This artificial clock displays noisy behaviour, possibly because of stochastic fluctuations of its components. Such 'rational network design may lead both to the engineering of new cellular behaviours and to an improved understanding of naturally occurring networks.	Curated		
		Modelling approach(es) ordinary differential equation model		
	The model is based upon the equations in Box 1 of the paper; however, these equations as printed are dimensionless, and the correct dimensions have been returned to the equations, and the parameters set to reproduce Figure 1C (left).	Connected external resources		
	The original model was generated by B.E. Shapiro using Cellerator version 1.0 update 2.1127 using Mathematica 4.2 for Mac OS X (June 4, 2002), November 27, 2002 12:15:32, using (PowerMac,PowerPC, Mac OS X,MacOSX,Darwin).	SBGN view in Newt Editor		
	Nicolas Le Novere provided a corrected version generated by SBMLeditor on Sun Aug 20 00:44:05 BST 2006. This removed the EmptySet species. Ran fine on COPASI 4.0 build 18.			
	Bruce Shapiro revised the model with SBMLeditor on 23 October 2006 20:39 PST. This defines default units and correct reactions. The original Cellerator reactions while being mathematically correct did not accurately reflect the intent of the authors. The original notes were mostly removed because they were mostly incorrect in the revised version. Tested with MathSBML 2.6.0.			
	Nicolas Le Novere changed the volume to 1 cubic micrometre, to allow for stochastic simulation.			



System Biology Models



Adapted from Elowitz and Leibler (2000) DOI: 10.1038/35002125



Taken from BioModels https://www.ebi.ac.uk/biomodels/ BIOMD0000000012#Curation

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The pipeline



ODE Models in SBML



```
</functionDefinition>
</listOfFunctionDefinitions>
<listOfUnitDefinitions>
 <unitDefinition id="length" name="length">
   <listOfUnits>
     <unit exponent="1" kind="metre" multiplier="1" scale="0"/>
   </listOfUnits>
 </unitDefinition>
 <unitDefinition id="area" name="area">
   <listOfUnits>
     <unit exponent="2" kind="metre" multiplier="1" scale="0"/>
   </listOfUnits>
 </unitDefinition>
 <unitDefinition id="volume" name="volume">
   <listOfUnits>
     <unit exponent="1" kind="litre" multiplier="1" scale="-3"/>
   </listOfUnits>
 </unitDefinition>
 <unitDefinition id="time" name="time">
   <listOfUnits>
     <unit exponent="1" kind="second" multiplier="86400" scale="0"/>
   </listOfUnits>
 </unitDefinition>
 <unitDefinition id="substance" name="substance">
```

<listOfUnits>



Import BioModels into odin



https://www.ebi.ac.uk/biomodels/BIOMD000000982

Import BioModels into odin

SIR model example (Malaysia)

importSBMLfromBioModels("BIOMD0000000982",".../LawModel.R")

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What is Menelmacar?

Menelmacar - Making Execution of (Nearly) Every Life-science Model ACcessible to All Researchers, helps to visualise models from the EMBL-EBI's BioModels database. You can investigate the models' development over time, change parameter values and observe changes in the model trajectory, and see the model graph.

Q. Enter model ID or name

Example models: BIOMD000000012 BIOMD000000003





Frequently asked questions

- Y What is this website for?
- ~ Who is this website for?
- How can I visualise a specific model?
- Y How do I know which models are available and how can I find models for a specific topic or organism?
- What information can be found in the different tabs of the website?

Noticed an error with a model? Report it here.

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Project idea and SBMLtoOdin: Leonie Lorenz and John Lees,





Code availability

- SBMLtoOdin: https://github.com/bacpop/SBMLtoOdin
- Menelmacar website:
 - Code: <u>https://github.com/bacpop/odinviewer</u>
 - Website: <u>biomodels.bacpop.org</u>
- NFDS Model: https://github.com/bacpop/compartmental-models-odin-dust



THANK YOU

Pathogen Informatics and Modelling Group

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